Andersen Lab Washing Protocols

Plastic and glass bottles, flasks, and beakers
1. Check the dirty area by the sink for any dishes that need to be washes

2. For each item:
   a. Fill up with regular tap water, swirl water around in container, and dump water
   b. Fill up with regular tap water and add a squirt of 10% Alconox soap
   c. Swirl soapy water around in container
   d. Rinse soapy water out with regular tap water until there are no more suds
   e. Rinse with deionized water (from white tap)

3. Lay each item to dry in the clean dishes area and let air dry

4. Prepare any glass bottles/flasks for autoclaving once they are dry

Stir Bars, Spatulas, and Scoops
1. Take pan of dirty spatulas, stir bars, and scoops to the sink.

2. For each item, do the following:
   a. Rinse in regular water.
   b. Rinse in distilled water (from the distilled water tap).
   c. Lay the item on a paper towel-lined tray.

3. Spray all items with 70% ethanol.

4. Wipe items off with a kim-wipe and return to their storage location.

5. Return the pan to hold the dirty spatulas, stir bars, and scoops to the chemical area.

Test Tubes
1. The tubes are in a tub of soapy water.

2. For each tube and cap, do the following:
   a. Scrub the tubes with a test tube brush and rinse out the soapy water.
   b. Squirt with 95% ethanol.
   c. Scrub using a test tube brush.
   d. Rinse with with regular $\text{H}_2\text{O}$.
   e. Rinse with distilled $\text{H}_2\text{O}$ 2-3 times.
f. Cap the tube and place it in a clean rack

3. Put a piece of autoclave tape on the rack.

4. Autoclave:
   a. Between the hours of 9 AM and 4 PM, autoclave in Pancoe 3343 on dry cycle for 10 minutes.
   b. After hours (before 9 AM or after 5 PM), autoclave on dry cycle for 10 minutes.

Glass Pipettes
1. Add a sprinkle of alconox soap (located under regular sink) to the first washing jar located in hood #2.
2. Fill the can with water up to the black line.

3. Transfer the pipette holder with dirty pipettes to the first washing pipette.
4. Allow the pipettes to soak for 10 minutes.

5. Turn on the regular water using the green knob outside the hood to allow the washing jar to fill and drain **five** times. If there are still suds after draining five times, fill and drain repeatedly until there are no more suds.

6. Transfer the pipette holder to the second washing can, attached to the distilled H$_2$O tap.

7. Turn on the tap from the orange knob and allow the washing can to fill and drain **three** times.

8. While the pipettes are rinsing, make sure there are paper towels in the bottom of the drying bin.

9. While wearing gloves, remove the pipettes and lay them in the drying bin to dry.

**96-well Plates For Rinsing the ReflX on the Worm Sorter**

1. Soak plates for 10 minutes in a tub of deionized water (white tap). Make sure to submerge the plates under water to remove all air bubbles in the wells.
2. Remove water from each plate by quickly flicking it into the sink. Dump out water from tub, rinse tub, and repeat Step 1 with fresh water.

3. Remove all water from each plate by quickly flicking it into the sink. Tap the remaining liquid out of the plates and allow them to dry upright in the large drying bin below the dish area.

4. Once dry, stack the plates WELL SIDE DOWN in the Clean Reflux plates bin by the sorter bench (this prevent dust particles from settling in the wells before use).

(This protocol was changed April 2014 per ECA to minimize sorter clogging - the former protocol is archived).